

the Science of the **Total Environment**

The Science of the Total Environment 248 (2000) 123-133

www.elsevier.com/locate/scitotenv

Occurrence of sulfonylurea, sulfonamide, imidazolinone, and other herbicides in rivers, reservoirs and ground water in the Midwestern United States, 1998

W.A. Battaglin^{a,*}, E.T. Furlong^b, M.R. Burkhardt^b, C.J. Peter^c

^aU.S. Geological Survey, Office of the Regional Hydrologist, Box 25046 MS 406, D.F.C., Denver, CO 80225, USA ^bU.S. Geological Survey, Methods Research and Development, Box 25046 MS 407, D.F.C., Denver, CO 80225, USA ^cDuPont Agricultural Products, Barley Mill Plaza, P.O. Box 80015, Wilmington, DE 19880-0015, USA

Received 5 April 1999; accepted 25 May 1999

Abstract

Sulfonylurea (SU), sulfonamide (SA), and imidazolinone (IMI) herbicides are relatively new classes of chemical compounds that function by inhibiting the action of a plant enzyme, stopping plant growth, and eventually killing the plant. These compounds generally have low mammalian toxicity, but plants demonstrate a wide range in sensitivity to SUs, SAs, and IMIs with over a 10 000-fold difference in observed toxicity levels for some compounds. SUs, SAs, and IMIs are applied either pre- or post-emergence to crops commonly at 1/50th or less of the rate of other herbicides. Little is known about their occurrence, fate, or transport in surface water or ground water in the USA. To obtain information on the occurrence of SU, SA, and IMI herbicides in the Midwestern United States, 212 water samples were collected from 75 surface-water and 25 ground-water sites in 1998. These samples were analyzed for 16 SU, SA and IMI herbicides by USGS Methods Research and Development Program staff using high-performance liquid chromatography/mass spectrometry. Samples were also analyzed for 47 pesticides or pesticide degradation products. At least one of the 16 SUs, SAs or IMIs was detected above the method reporting limit (MRL) of 0.01 µg/l in 83% of 130 stream samples. Imazethapyr was detected most frequently (71% of samples) followed by flumetsulam (63% of samples) and nicosulfuron (52% of samples). The sum of SU, SA and IMI concentrations exceeded 0.5 μg/l in less than 10% of stream samples. Acetochlor, alachlor, atrazine, cyanazine and metolachlor were all detected in 90% or more of 129 stream samples. The sum of the concentration of these five herbicides exceeded 50 µg/l in

*Corresponding author. Tel.: +1-303-236-5950, ext. 202; fax: +1-303-235-5919. E-mail address: wbattagl@usgs.gov (W.A. Battaglin)

0048-9697/00/\$ - see front matter © 2000 Elsevier Science B.V. All rights reserved.

PII: S0048-9697(99)00536-7

approximately 10% of stream samples. At least one SU, SA, or IMI herbicide was detected above the MRL in 24% of 25 ground-water samples and 86% of seven reservoir samples. © 2000 Elsevier Science B.V. All rights reserved.

Keywords: Sulfonylurea; Sulfonamide; Imidazolinone; Herbicides; Rivers; Reservoirs; Ground water

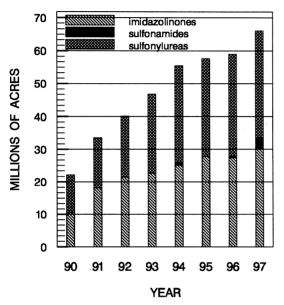


Fig. 1. Estimated acres of corn, soybeans and wheat treated with selected SU, SA, and IMI herbicides, 1990–1997, in 11 Midwestern United States.

1. Introduction

During the last 20 years, low application rate herbicides have been developed that act by inhibiting the action of a key plant enzyme, which stops plant growth and eventually causes plant death. Sulfonylurea (SU), sulfonamide (SA), and imidazolinone (IMI) herbicides are three classes of compounds that share this mode of action (Meister, 1997).

1.1. Herbicide use

Crops that can be treated with SU, SA and IMI herbicides include barley, corn, cotton, durum wheat, rice, canola, peanuts, soybeans, sugar beets, spring wheat and winter wheat. Some compounds are also approved for use on Conservation Re-

serve Program acreage and for non-cropland weed control.

The amount of cropland treated with SU, SA, and IMI herbicides has nearly tripled since 1990. The total corn, soybean and wheat acreage on which nine SUs, one SA and two IMIs were applied in 11 Midwestern States (Iowa, Illinois, Indiana, Kansas, Kentucky, Minnesota, Missouri, Nebraska, Ohio, South Dakota and Wisconsin) from 1990 to 1997 is shown in Fig. 1 (US Department of Agriculture, 1991–1998). In 1997, more than 66 000 000 acres were treated with one of the 12 herbicides. For comparison, atrazine, a triazine herbicide, was used on 41 000 000 acres of corn and metolachlor, a chloracetanilide herbicide, was used on 23 000 000 acres of corn and soybeans in the same 11-state area.

Although applied over comparable areas, SU, SA and IMI herbicides are frequently applied after crops have emerged, and at low rates (typically less than 25 g active ingredient/ha). These application rates are commonly 1/50th or less of the rates for triazine or chloracetanilide herbicides (typically more than 1200 g/ha). Hence, the overall use amount for SU, SA and IMI herbicides is relatively small. For example, in 1997 in the 11-State area, an estimated 20 300 t of atrazine and 21 500 t of metolachlor were applied to cropland, while the total estimated use of the nine SUs, one SA, and two IMIs was only 1090 t (US Department of Agriculture, 1991–1998).

1.2. Herbicide chemistry

The soil half-life of SUs, SAs, and IMIs generally ranges from 1 to 25 weeks depending on soil pH and temperature. Their water solubilities range from 6 to 40 000 part per million. The water solubility of SUs is dependent on water pH. SUs degrade by chemical hydrolysis and microbial ac-

tivity. SUs degrade faster in warm, moist, low organic, low pH soils (DuPont, 1998). IMIs degrade by microbial activity and photolysis. IMIs degradate faster in warm, moist, low organic soils (Goetz et al., 1990).

1.3. Herbicide toxicity

SUs, SAs and IMIs act upon a specific plant enzyme (acetolactate synthase) that is not found in mammals or other animals and are reported to have very low toxicities in animals (Brown, 1990: Meister, 1997). Plants demonstrate a wide range in sensitivity to SUs, SAs and IMIs (Peterson et al., 1994) with over a 10000-fold difference in observed toxicity levels for some compounds. EC₅₀ concentrations are measures of compound toxicity. An EC₅₀ is the concentration in water of a compound that causes a 50% reduction in a chosen plant characteristic for which a toxicity endpoint exists. For example, EC₅₀ values for algae can be calculated from laboratory tests measuring biomass development in the presence of varying compound concentrations. EC₅₀ values for selected SU, SA, IMI and other herbicides on five aquatic plants are shown in Fig. 2 (Fahl et al., 1995; Fairchild et al., 1997; Sabater and Carrasco, 1997; Wei et al., 1998; US Environmental Protection Agency, 1997; Peter, DuPont Agricultural Products, written communication, 1999). The EC₅₀ values plotted are for green algae (Selenastrum capricornutum), duckweed (Lemna gibba), blue-green algae (Anabaena flos-aquae), freshwater algae (Scenedesmus costatum), and freshwater diatom (*Navicula pelliculosa*). In some cases, EC₅₀ values from more than one test on the same plant species are included. EC₅₀ values for several herbicides range over 3 orders of magnitude. The EC₅₀ data plotted in Fig. 2 support the hypothesis that a concentration of $0.1 \mu g/1$ in water is the baseline for non-target aquatic plant toxicity.

SUs, SAs, and IMIs are active at very low concentrations and can cause a problem with plant vigor in some crop rotations even when only 1% or less of the originally applied material remains. Some of these herbicides have demonstrated residual phytotoxicity to rotation crops such as corn, sunflowers, sugar beets and dry

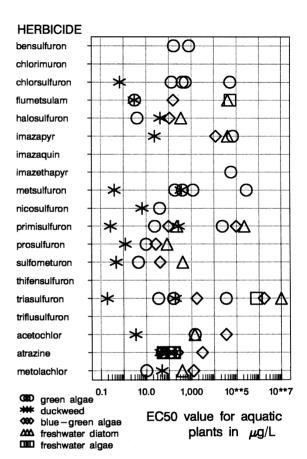


Fig. 2. EC₅₀ concentrations in micrograms per liter for five aquatic plants for selected SU, SA, IMI and other herbicides.

beans (Anderson and Humburg, 1987; Curran et al., 1991). The labels of some of these herbicides restrict the planting of certain rotational crops. Fletcher et al. (1993) indicated that spray drift containing SUs at concentrations less than 1% of the recommended application rate may adversely impact fruit tree yields. Felsot et al. (1996) suggested that the appearance of chlorotic spots on crops in south central Washington is a result of exposure to low levels of SU herbicides in precipitation and not from direct spray drift. However, Obrigawitch et al. (1998) questioned the validity of Fletcher's findings and the results of other studies that based their findings on short-term plant-response assessments. Obrigawitch et al. (1998) found that a treatment rate of 0.1 g of the most active SU ingredient per hectare (0.00009 lb

per acre) represents a 'threshold dose' and would be unlikely to reduce the yields of even the most sensitive non-target plants.

1.4. Herbicide occurrence

Detections of SUs, SAs and IMIs in water collected from environmental settings have been rare and the few reported detections have been at nanogram per liter concentrations (Bergstrom, 1990; Michael and Neary, 1993; D'Ascenzo et al., 1998; Okamoto et al., 1998; Steinheimer et al., 1999). However, several studies indicate that some SUs, SAs and IMIs herbicides may leach beyond the active root zone and enter ground-water or surface-water systems (Anderson and Humburg, 1987; Bergstrom, 1990; Veeh et al., 1994; Flury et al., 1995). Once in ground water or surface water, some SUs, SAs and IMIs will tend to persist as the parent compound while others will tend to hydrolyze (Harvey et al., 1985; Dinelli et al., 1997). A study by Afyuni et al. (1997) indicated that between 1.1 and 2.3% of an applied SU was lost in runoff during a simulated rainfall event 24 h after herbicide application.

Because of their low application rates and low overall use amounts, concentrations of SUs, SAs and IMIs are expected to be low in most water resources. One can also assume, based upon their chemical characteristics, application rates, and acres treated that individual SUs, SAs and IMIs herbicides would be expected to occur in surface or ground water at 1-0.1% or less of the concentration of common triazine herbicides. Thus, one could expect to observe SUs, SAs and IMIs herbicides in Midwestern rivers during post-application runoff events at concentrations ranging from 0.001 to 0.1 µg/l. Furthermore, one could expect maximum concentrations of SUs, SAs and IMIs herbicides to range from 0.01 to 1.0 µg/l (Battaglin et al., 1998a). The concentrations of triazine herbicides observed in ground water (Kolpin et al., 1994) are generally 1-2 orders of magnitude less than those observed in streams during post-application runoff. Hence, one would expect SU, SA and IMI concentrations to seldom exceed 0.01 μg/l in ground water.

2. Objectives and methods

2.1. Purpose and scope

Currently, little is known about the occurrence, fate, or transport of SUs, SAs and IMIs in the hydrologic system in the USA. The overall objective of this project is to determine if and at what concentrations selected SUs, SAs, and IMIs occur in surface- and ground-water resources of the Midwestern USA. Specific objectives include:

- Develop an analytical method for selected SUs. SAs and IMIs.
- Conduct a reconnaissance to determine the environmental occurrence of SUs, SAs and IMIs herbicides in surface water and ground water in the Midwestern USA.
- Determine the frequency of detection and concentration distributions of SUs, SAs and IMIs herbicides relative to those of selected other herbicides in midwestern surface water and ground water.

A Cooperative Research and Development Agreement (CRADA) between the US Geological Survey (USGS) and DuPont Agricultural Products was developed to accomplish the objectives of this project in an unbiased, yet economical manner (Battaglin et al., 1998b).

The data collected in this study are only adequate to identify the occurrence of selected SU, SA and IMI herbicides during post-application runoff events in Midwestern streams and in ground water in parts of Iowa and Illinois. The data are not adequate to determine annual mean concentrations of detected herbicides or whether non-detected herbicide are present at other times of the year.

2.2. Plan of study

The study involved the collection of 212 samples during a 1998 reconnaissance. Samples were collected from streams, large rivers, reservoir outflows, and wells, sometimes in conjunction with USGS National Stream Quality Accounting Network (NASQAN) (Hooper et al., 1997) and Natio-

Table 1 Statistical summary of SU, SA, IMI, herbicide concentrations in 130 samples and selected other herbicide concentrations in 129 water samples from Midwestern streams and rivers, 1998 (in $\mu g/l$)

Herbicide	Detections above MRL	Median	Maximum
SU, SA and IMI herbicides			
Bensulfuron methyl	0	< 0.01	< 0.01
Chlorimuron ethyl	41	< 0.01	0.304
Chlorsulfuron	1	< 0.01	0.013
Flumetsulam	82	0.020	0.358
Halosulfuron methyl	7	< 0.01	0.067
Imazapyr	5	< 0.01	0.072
Imazaquin	41	< 0.01	1.11
Imazethapyr	92	0.031	0.689
Metsulfuron methyl	0	< 0.01	< 0.01
Nicosulfuron	67	0.010	0.266
Primisulfuron methyl	0	< 0.01	< 0.01
Prosulfuron	6	< 0.01	0.036
Sulfometuron methyl	2	< 0.01	0.018
Thifensulfuron methyl	1	< 0.01	0.015
Triasulfuron	0	< 0.01	< 0.01
Triflusulfuron methyl	0	< 0.01	< 0.01
Sum of 16 SUs, SAs and IMIs	108	0.137	2.10
Other herbicides			
Acetochlor	124	0.411	25.1
Alachlor	116	0.045	17.2
Atrazine	129	3.97	224.
Cyanazine	119	0.326	14.0
Metolachlor	129	1.73	143.
Sum of five other herbicides	129	6.90	385.

nal Water Quality Assessment (NAWQA) (Leahy and Thompson, 1994) activities. All reconnaissance samples were analyzed for 16 SU, SA and IMI herbicides (Table 1) using high performance liquid chromatography coupled with mass spectrometry. This custom analytical method has a method reporting limit (MRL) of $0.01 \,\mu\text{g/l}$ for all analytes and is fully described in Furlong et al. (1999).

2.3. Sampling sites

Samples were collected from 75 surface-water sites in the Upper Mississippi, Missouri, and Ohio River basins (Fig. 3). Fifty-two of the surface-water

sites have been studied in previous Midcontinent Herbicide Initiative (MHI) investigations (Thurman et al., 1992; Goolsby et al., 1994; Scribner et al., 1998). These sites were selected out of the set of 150 sites sampled in 1989 using a stratified random method (Scribner et al., 1993). It is important to note that the sampling strategy used was not designed to produce an unbiased estimate of herbicide occurrence in all midwestern streams. Rather the intent was to target higher risk areas while still capturing the variability of the entire population. Samples also were collected at selected NASOAN and NAWOA sites and just downstream from five reservoirs at locations that had been sampled in a previous investigation (Coupe et al., 1995; Scribner et al., 1996).

Twenty ground-water samples were collected from a network of municipal wells in Iowa that are part of the Iowa Ground water Monitoring (IGWM) program (Detroy et al., 1988; Kolpin et al., 1997). Wells from this network have been sampled systematically since 1982. The depths to the top of the well screen for the 20 sampled wells, ranged from 6 to 83 m with 16 of the 20 being less than 20 m. Samples also were collected from five observation wells in the Lower Illinois

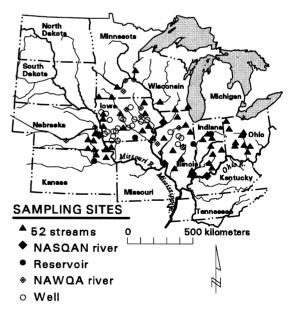


Fig. 3. Location and type of sites sampled in 1998 herbicide reconnaissance.

NAWQA study unit (Warner and Schmidt, 1994). These wells were all less than 8 m deep.

2.4. Sampling schedule and procedure

Two samples were collected at each surfacewater and reservoir site, and one sample was collected at each ground water site in 1998. The first surface-water samples were collected after pre-emergence herbicides were applied (May or June) and following a precipitation event that produced a significant increase in streamflow. These samples will be referred to as pre-emergence runoff samples. The second surface-water samples were collected after post-emergence herbicides were applied (June or July) again following a precipitation event that produced runoff and an increase in streamflow. These samples will be referred to as post-emergence runoff samples. The first NASQAN and reservoir samples were collected 2-3 weeks after the first surface-water samples were collected from nearby sites. The second NASQAN and reservoir samples were collected 2-3 weeks after the second surface-water samples were collected from nearby sites. Ground-water samples were collected in June, July, or August.

Samples were collected using protocols that are identical to those used for the collection of samples for low levels of other dissolved organic compounds (Shelton, 1994). The equal-width-increment sampling method was used in all cases except on some large rivers where equal-discharge-increment sampling was used (Edwards and Glysson, 1988).

All equipment was precleaned with a Liquinox/tap-water solution, rinsed with tap water, deionized water, and then methanol, and air dried. All samples were filtered through 0.7-µm poresize heat-cleaned glass-fiber filters using an aluminum-plate filter holder and a ceramic-piston fluid-metering pump with all Teflon tubing into precleaned 1-l or 125-ml amber glass bottles. Samples were immediately chilled and shipped on ice from the field to the USGS National Water Quality Laboratory (NWQL) within 2 days of collection.

2.5. Analytical methods

An analytical method was developed that is an extension and improvement of the high-performance liquid chromatography/mass spectrometry (HPLC/MS) method by Rodriguez and Orescan (1998). Briefly, the method consists of isolation of the analytes of interest from approximately 1 l of water (precisely measured) using two stacked solid-phase extraction cartridges. After isolation, the second cartridge is dried and the analytes are eluted using acidified acetone. The analytes are then concentrated and solvent exchanged into 1 ml of 10% acetonitrile and 90% water.

HPLC/MS analysis is performed using a Hewlett Packard 1100 series HPLC, coupled to a Hewlett Packard LC/MSD. Electrospray ionization, operated in the positive ion mode, is used to ionize the analytes. Selected-ion monitoring is used to maximize sensitivity. A calibration curve is developed using external standards and the linear range of the method is from 0.005 to 0.5 µg/l. Three ions (one quantitation, two confirmation) are monitored for each compound. For confirmed identification of analytes, the relative ion abundances for the detection must be within 20% of the average response for standards, as well as have the correct relative retention time (within 0.1 min). Detected analytes that don't meet one criterion are reported as estimates, as are all detections below the MRL of 0.01 µg/l. Details of this analytical methods are provided elsewhere in this volume by Furlong et al. (1999).

In addition, all samples were analyzed for several other classes of pesticides. Samples were analyzed for 41 pesticides and pesticide metabolites by GC/MS with selected-ion monitoring using methods described by Zaugg et al. (1995). This method has MRLs that range from 0.001 to $0.018~\mu g/l$.

3. Results and discussion

Sample collection began in May and was completed in August, 1998. Two hundred and twelve samples were submitted to Methods Research and Development Program staff at the NWQL.

Of these, 177 are from surface-water sites and 29 are from ground water sites (results from six samples collected from two tile drains in New York State are not discussed here). These numbers include QA samples. One hundred and fifty surface-water samples and 25 ground-water samples were submitted to the NWQL for analysis of 47 pesticides or pesticide degradation products.

3.1. Occurrence of SU, SA and IMI herbicides in surface water

Results from 130 stream and river samples are summarized in Table 1. At least one of the 16 SU. SA or IMI herbicides was detected in 83% of stream samples. Imazethapyr was the most frequently detected compound (71% of samples) followed by flumetsulam (63%), nicosulfuron (52%), imazaquin and chlorimuron ethyl (32%). Chlorsulfuron, halosulfuron methyl, imazapyr, prosulfuron, sulfometuron methyl and thifensulfuron methyl were detected in 5% or less of samples. The detection frequencies reported are for samples with concentrations at or above the method reporting limit (MRL), currently 0.01 µg/l. Bensulfuron methyl, metsulfuron methyl, primisulfuron methyl, triasulfuron, and triflusulfuron methyl were not detected above the MRL.

The distributions of concentrations of the target analytes in 130 samples are summarized in Fig. 4. In some cases, estimated concentrations are reported that are below the MRL. These concentrations are not counted as detections above the MRL (numbers to the right of the boxplots in Fig. 4), but are used in the calculation of summary statistics (the boxplots themselves). The sum of SU, SA, and IMI concentrations exceeded 0.5 µg/l in less than 10% of stream samples. At least one of the 16 SU, SA or IMI herbicides was detected above the MRL in six of seven (86%) reservoir samples. Flumetsulam, imazethapyr, imazaquin, and nicosulfuron were each detected in four samples. The sum of SU, SA and IMI concentrations did not exceed 0.5 μg/l in any reservoir sample.

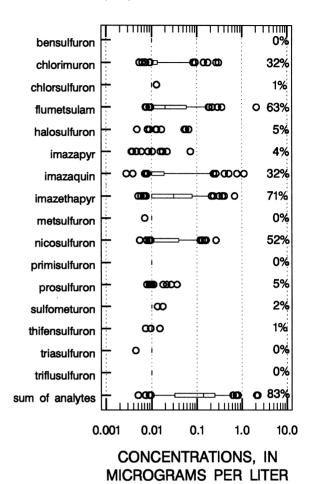


Fig. 4. Boxplots of SU, SA and IMI herbicide concentrations and percent detections above the MRL $(0.01~\mu g/l)$ in 130 samples from midwestern streams, 1998. The boxes show the 25th, 50th (median) and 75th percentiles, the wiskers extend to the 5th and 95th percentiles, and outliers less than the 5th or greater than the 95th percentiles are shown as circles.

3.2. Occurrence of other herbicides in surface water

The results of analysis for selected other herbicides in 129 stream or river samples are also included in Table 1. Acetochlor, alachlor, atrazine, cyanazine and metolachlor were all detected in 90% or more of the stream samples. Atrazine had the highest median concentration (3.97 μ g/l), followed by metolachlor (1.73 μ g/l), and acetochlor (0.411 μ g/l). The sum of the five other

herbicides included in Table 1 exceeded 50 $\mu g/l$ in approximately 10% of the samples. This sum was expected to be at least 100 times greater than the sum of the SU, SA and IMI herbicide concentrations.

3.3. Occurrence of SU, SA and IMI herbicides in ground water

Results from 25 ground-water samples are summarized in Table 2. At least one of the 16 SUs, SAs or IMIs was detected in 24% of ground-water samples. Imazethapyr was the most frequently detected compound (16% of samples) followed by flumetsulam (12%), and nicosulfuron and imazaquin (8%). The sum of SU, SA and IMI concentrations exceeded 0.01 µg/l in six samples.

3.4. Occurrence of other herbicides in ground water

The results of analysis for selected other herbicides are also included in Table 2. Atrazine and metolachlor were detected in approximately half of the samples. The sum of the concentrations of the five herbicides did not exceeded 1.0 μ g/l in any sample. This sum was also expected to be at least 100 times greater than the sum of the SU, SA and IMI herbicide concentrations.

4. Concentrations of SU, SA and IMI herbicides relative to other herbicides

Because they have similar chemical properties, but much lower application rates, and a shorter history of use, SU, SA and IMI herbicides were expected to occur at a fraction (1/100th or less) of the concentrations of other herbicides such as atrazine (Battaglin et al., 1998a). In Fig. 5a,b, and c the concentrations in streams of imazethapyr, flumetsulam and nicosulfuron, the three most frequently detected target analytes, are plotted vs. atrazine concentration. The circles are pre-emergence samples and the triangles are post-emergence samples. The lines crossing these plots show the 1:10, 1:100, and 1:1000 ratios of concentration. The data plotted on Fig. 5 suggest that in approximately half the samples, imazethapyr,

Table 2 Statistical summary of SU, SA, IMI and selected other herbicide concentrations in 25 samples of Midwestern ground water, 1998 (in μg/l)

Herbicide	Detection above MRL	Median	Maximum
SU, SA and IMI herbicides			
Bensulfuron methyl	0	< 0.01	< 0.01
Chlorimuron ethyl	0	< 0.01	< 0.01
Chlorsulfuron	0	< 0.01	< 0.01
Flumetsulam	3	< 0.01	0.035
Halosulfuron methyl	0	< 0.01	< 0.01
Imazapyr	0	< 0.01	< 0.01
Imazaquin	2	< 0.01	0.024
Imazethapyr	4	< 0.01	0.059
Metsulfuron methyl	0	< 0.01	< 0.01
Nicosulfuron	2	< 0.01	0.016
Primisulfuron methyl	0	< 0.01	< 0.01
Prosulfuron	0	< 0.01	< 0.01
Sulfometuron methyl	0	< 0.01	< 0.01
Thifensulfuron	0	< 0.01	< 0.01
Triasulfuron	0	< 0.01	< 0.01
Triflusulfuron methyl	0	< 0.01	< 0.01
Sum of 16 SUs, SAs, and IMIs	6	< 0.01	0.110
Other herbicides			
Acetochlor	1	< 0.002	0.004
Alachlor	1	< 0.002	0.016
Atrazine	14	0.010	0.410
Cyanazine	I	< 0.004	0.007
Metolachlor	12	< 0.002	0.557
Sum of five other herbicides	14	0.014	0.703

flumetsulam and nicosulfuron occur at 1/100th or less of the concentration of atrazine.

SU, SA and IMI concentrations were expected to be smaller relative to atrazine concentrations in the pre-emergence samples than in the post-emergence samples because the majority of atrazine is applied before crops emerge, and the majority of the SU, SA and IMI compounds are applied after crops emerge. In 68 pre-emergence runoff samples from Midwestern streams, the median atrazine to imazethapyr, flumetsulam, and nicosulfuron ratios were 146, 173 and 336, respectively. In 62 post-emergence runoff samples from Midwestern streams, the median atrazine to imazethapyr, flumetsulam, and nicosulfuron ratios were 48, 75 and 68, respectively.

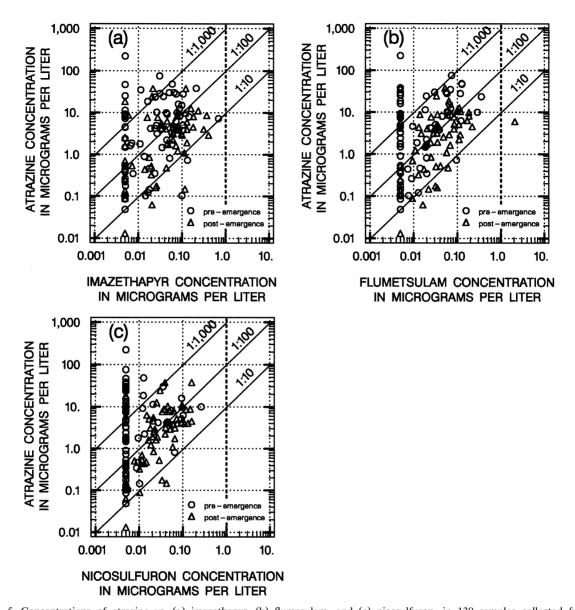


Fig. 5. Concentrations of atrazine vs. (a) imazethapyr, (b) flumesulam, and (c) nicosulfuron, in 130 samples collected from Midwestern streams in 1998.

The observed range and maximum concentrations of SU, SA and IMI herbicide in samples collected from Midwestern streams during postapplication runoff events in 1998 was very close to what we expected. The majority of SU, SA and IMI detections were at concentrations less than 0.1 µg/l. These concentrations are not likely to

be toxic to non-target aquatic plants nor are they likely to be of concern for human consumption, but they do add to the overall burden of pesticides carried by Midwestern rivers. The maximum concentrations of SU, SA and IMI herbicides in samples collected from midwestern ground water in 1998 were slightly higher than expected.

Acknowledgements

This research was conducted as part of the US Geological Survey's Toxic Substances Hydrology Program in cooperation with E.I. DuPont. The authors are grateful for reviews by Dana Kolpin, Donald Goolsby and John Flager, at the USGS; and two anonymous reviewers.

References

- Afyuni MM, Wagger MG, Leidy RB. Runoff of two sulfonylurea herbicides in relation to tillage system and rainfall intensity. J Environ Qual 1997;26:1318–1326.
- Anderson RL, Humburg NE. Field duration of chlorsulfuron bioactivity in the central great plains. J Environ Qual 1987;16:263–266.
- Battaglin WA, Furlong ET, Burkhardt M, Peter CJ. A reconnaissance for new, low-application rate herbicides in surface and ground water in the midwestern United States, 1998. In: Proceeding of the NWQMC National Conference Monitoring: Critical Foundations to Protect Our Waters US Environmental Protection Agency 1998:245–256.
- Battaglin WA, Furlong ET, Peter CJ. A reconnaissance for sulfonylurea herbicides in waters of the Midwestern USA: An example of collaboration between the public and private sectors. US Geol Surv Fact Sheet FS-046-98, 1998:4.
- Bergstrom L. Leaching of chlorsulfuron and metsulfuron methyl in three Swedish soils measured in field lysimeters. J Environ Qual 1990;19:701–706.
- Brown HM. Mode of action, crop selectivity, and soil relations of the sulfonylurea herbicide. Pestic Sci 1990;29:263–281.
- Coupe RH, Goolsby DA, Iverson JL, Markovchick DJ, Zaugg SJ. Pesticide, nutrient, water-discharge and physical-property data for the Mississippi River and some of its tributaries, April 1991–September 1992. US Geol Surv Open-File Rep 93–657, 1995:116.
- Curran WS, Knake EL, Liebl RA. Corn (*Zea mays*) injury following use of clomazone, chlorimuron, imazaquin, and imazethapyr. Weed Technol 1991;5:539–544.
- D'Ascenzo G, Gentili A, Marchese S, Marino A, Perret D. Multiresidue method for determination of post-emergence herbicides in water by HPLC/ESI/MS in positive ionization mode. Environ Sci Technol 1998;32:1340–1347.
- Detroy MG, Hunt PKB, Holub MA. Ground-water-qualitymonitoring program in Iowa: nitrate and pesticides in shallow aquifers. US Geol Surv Open-File Rep 88–4123, 1988:32.
- Dinelli G, Vicari A, Bonetti A, Catizone P. Hydrolytic dissipation of four sulfonylurea herbicides. J Agric Food Chem 1997;45:1940–1945.
- DuPont. A guide to DuPont sulfonylurea herbicides. Wilmington, DE: E.I. du Pont de Nemours and Company, 1998.
- Edwards TK, Glysson DG. Field methods for measurement of

- fluvial sediment. US Geol Surv Open-File Rep 86-531, 1988:118.
- Fairchild JF, Ruessler DS, Haverland PS, Carlson AS. Comparative sensitivity of *Selenastrum capricomutum* and *Lemma minor* to sixteen herbicides. Arch Environ Contam Toxicol 1997;32:353–357.
- Fahl GM, Kreft L, Altenburger R, Faust M, Boedeker W, Grimme LH. pH-dependent sorption, bioconcentration and algal toxicity of sulfonylurea herbicides. Aquat Toxicol 1995;31:175–187.
- Felsot AS, Bhatti MA, Mink GI, Reisenauer G. Biomonitoring with sentinel plants to assess exposure of nontarget crops to atmospheric deposition of herbicide residues. Environ Toxicol Chem 1996;15:452–459.
- Fletcher JS, Pfleeger TG, Ratsch HC. Potential environmental risks associated with the new sulfonylurea herbicides. Environ Sci Technol 1993;27:2250–2252.
- Flury M, Leuenberger J, Studer B, Fluhler H. Transport of anions and herbicides in a loamy and a sandy field soil. Water Resour Res 1995;31:823–835.
- Furlong ET, Burkhardt MR, Gates PM, Werner MG, Battaglin WA. Routine determination of sulfonylurea, imidazolinone, and sulfonamide herbicide at parts per trillion concentrations by solid-phase extraction and liquid chromatography/mass spectrometry. Sci Total Environ 1999: this volume.
- Goetz AJ, Lavy TL, Gbur EE. Degradation and field persistence of imazethapyr. Weed Sci 1990;38:421–428.
- Goolsby DA, Boyer LL, Battaglin WA. Plan of study to determine the effect of changes in herbicide use on herbicide concentrations in midwestern streams, 1989–94. US Geol Surv Open-File Rep 94–347, 1994:14.
- Harvey J, Dulka JJ, Anderson JJ. Properties of sulfometuron methyl affecting its environmental fate: Aqueous hydrolysis and photolysis, mobility and adsorption on soils, and bioaccumulation potential. J Agric Food Chem 1985;33:590–596.
- Hooper RP, Goolsby DA, Rickert DA, McKenzie SW. NASQAN a program to monitor the water quality of the Nation's large rivers. US Geol Surv Fact Sheet FS-055-97, 1997:6.
- Kolpin DW, Burkart, MR, Thurman EM. Herbicides and nitrate in near-surface aquifers in the midcontinental United States, 1991. US Geol Surv Water-Supply Paper 2413, 1994:34.
- Kolpin DW, Kalkhoff SJ, Goolsby DA, Sneck-Fahrer DA, Thurman EM. Occurrence of selected herbicides and herbicide degradation products in Iowa's ground water, 1995. Groundwater 1997;35:679–688.
- Leahy PP, Thompson TH. The National Water Quality Assessment Program. US Geol Surv Open-File Rep 94–70, 1994.
- Meister RT. Farm chemicals handbook '97. Willoughby, Ohio: Meister Publishing Company, 1997: various pagination.
- Michael JL, Neary DG. Herbicide dissipation studies in southern forest ecosystems. Environ Toxicol Chem 1993;12: 405-410.

- Obrigawitch TT, Cook G, Wetherington J. Assessment of effects on non-target plants from sulfonylurea herbicides using field approaches. Pestic Sci 1998;52:199–217.
- Okamoto Y, Fisher RL, Armbrust KL, Peter CJ. Surface water monitoring survey for bensulfuron methyl applied in paddy fields. Pestic Sci 1998;23:235–240.
- Peterson HG, Boutin C, Martin PA, Freemark KE, Ruecker NJ, Moody MJ. Aquatic phyto-toxicity of 23 pesticides applied at expected environmental concentrations. Aquat Toxicol 1994;28:275–292.
- Rodriguez M, Orescan DB. Confirmation and quantitation of selected sulfonylurea, imidazolinone, and sulfonamide herbicides in surface water using electrospray LC/MS. Anal Chem 1998;70:2710–2717.
- Sabater C, Carrasco JM. Effects of chlorsulfuron on growth of three freshwater species of phytoplankton. Bull Environ Contam Toxicol 1997;58:807–813.
- Scribner EA, Thurman EM, Goolsby DA, Meyer MT, Mills MS, Pomes ML. Reconnaissance data for selected herbicides, two atrazine metabolites, and nitrate in surface water of the Midwestern United States, 1989–1990. US Geol Surv Open-File Rep 93–457, 1993:77.
- Scribner EA, Goolsby DA, Thurman EM, Meyer MT, Battaglin WA. Concentrations of selected herbicides, herbicide metabolites, and nutrients in outflow from selected midwestern reservoirs, April 1992 through September 1993. US Geol Surv Open-File Rep 96–393, 1996:128.
- Scribner EA, Goolsby DA, Thurman EM, Battaglin WA. A reconnaissance for selected herbicides, metabolites, and nutrients in streams of nine Midwestern States, 1994–95. US Geol Surv Open-File Rep 98–181, 1998:44.
- Shelton LR. Field guide for collection and processing streamwater samples for the national water-quality assessment program. US Geol Surv Open-File Rep 94–455, 1994:42.
- Steinheimer TR, Pfeiffer RL, Peter CJ, Duffy MJ, Battaglin

- WA. Reconnaissance survey of sulfonamide, sulfonylurea, and imidazolinone herbicides in surface streams and groundwater of the Midwestern US. In: Steinheimer TR, Ross LJ, Spittler TD, editors. Agrochemical transport: perspective and scale-of study. American Chemical Society, Symposium Series, December, 1999.
- Thurman EM, Goolsby DA, Meyer MT, Kolpin DW. A reconnaissance study of herbicides and their metabolites in surface water of the Midwestern United States using immunoassay and gas chromatography/mass spectrometry. Environ Sci Technol 1992;26:2440–2447.
- US Department of Agriculture. Agricultural chemical usage: 1990–1997 field crops summary. Washington, DC: US Depart of Agriculture, NASS, ERS, 1991–1998: published annually.
- US Environmental Protection Agency. US Environmental Protection Agency, Office of Pesticide Programs, Environmental Fate and Effects Division, Tox One-Liner Database, 1997
- Veeh RH, Inskeep WP, Roe FL, Ferguson AH. Transport of chlorsulfuron through soil columns. J Environ Qual 1994; 23:542-549.
- Warner KL, Schmidt AR. National water-quality assessment program — the lower Illinois River basin. US Geol Surv Fact Sheet FS-94-018, 1994:2.
- Wei L, Yu H, Sun Y, Fen J, Wang L. The effects of three sulfonylurea herbicides and their degradation products on the green algae *Chlorella pyrenoidosa*. Chemosphere 1998;37:747–751.
- Zaugg SD, Sandstrom MW, Smith SG, Fehlberg KM. Methods of analysis by the U.S. Geological Survey National Water Quality Laboratory determination of pesticides in water by C-18 solid-phase extraction and capillary-column gas chromatography/mass spectrometry with selected-ion monitoring. US Geol Surv Open-File Rep 95–181, 1995:49.